








# From gene to table – DNA barcoding the backbone of next-generation food integrity and safety

Aleksandra Martinović<sup>1\*</sup> , Nađa Raičević , Beatriz Daza Prieto<sup>2</sup> , Werner Ruppitsch<sup>1,3</sup>  and Marija Vugdelić<sup>1</sup> 

<sup>1</sup> University of Donja Gorica, Faculty for Food Technology, Food Safety and Ecology, Oktoih 1, 81000 Podgorica, Montenegro

<sup>2</sup> Institute of Medical Microbiology and Hygiene, Austrian Agency for Health and Food Safety, Vienna, Austria

<sup>3</sup> Medical University Innsbruck, Institute of Hygiene and Medical Microbiology, Schöpfstraße 41, 6020 Innsbruck, Austria

## ARTICLE INFO

### Keywords:

DNA barcoding  
Food authenticity  
Food safety  
Molecular identification  
Traceability  
Food fraud

## ABSTRACT

The authenticity and safety of food are critical issues in global supply chains, where mislabelling, adulteration and contamination continue to challenge food integrity, consumer-trust and safety, and public health. DNA barcoding, a molecular identification method based on short genetic sequences, is a powerful tool for species verification in a wide range of food products, including meat, fish, plant-based foods, and processed foods. This study explores the application of DNA barcoding with an emphasis on its role against food fraud, consumer protection and public health. Advances in barcoding technologies, coupled with accessible databases enable identification of species even in complex or degraded samples. Case examples illustrate its successful implementation in regulatory, industrial, and research settings. The integration of DNA barcoding into traceability systems supports a more transparent, accountable, and trustworthy food supply industry. As food systems evolve, DNA-based verification is poised to become a core component of next-generation food integrity strategies.

## 1. Introduction

Food quality and safety are critical issues in the increasing global food trade characterized by complex supply chains. Although consumers increasingly demand sustainable, transparent, and healthy food products — and despite stricter regulations and technological progress — food fraud remains a persistent issue. It poses risks to public health, causes economic damage, compromises religious dietary requirements, and may even contribute to the endangerment of protected species (Li *et al.*, 2020; Filonzi *et al.*, 2023). Food fraud involves deliberate mislabelling, species substitution, and exposure to toxins,

pathogens and allergens (Ortea *et al.*, 2012) threatening consumer health and undermine consumer trust (Dawan and Ahn, 2022). Mislabelling rates across various food sectors including seafood, meat, botanicals, spices, and probiotics exceed on average by > 20% (Gorini *et al.*, 2023) to a maximum of >70% for certain food categories (Stamatis *et al.*, 2015). The most common type of food fraud is the replacement of a component with a similar cheaper one, followed by undeclared ingredients (Stamatis *et al.*, 2015).

Since the 1980s worldwide research strategies have been aiming to develop and improve

\*Corresponding author: Aleksandra Martinović, [aleksandra.martinovic@udg.edu.me](mailto:aleksandra.martinovic@udg.edu.me)

Paper received September 10<sup>th</sup> 2025. Paper accepted September 15<sup>th</sup> 2025.

The paper was presented at the 63<sup>rd</sup> International Meat Industry Conference “Food for Thought: Innovations in Food and Nutrition” – Zlatibor, October 05<sup>th</sup>–08<sup>th</sup> 2025.

Published by Institute of Meat Hygiene and Technology – Belgrade, Serbia.

This is an open access article CC BY licence (<http://creativecommons.org/licenses/by/4.0>)

technologies to control authenticity and safety of foods (Katerinopoulou et al., 2020). Amongst the analytical tests used for food authentication DNA-based techniques are most effective. DNA barcoding, a molecular technique that uses short, standardized genetic markers to identify species (Hebert et al., 2003), has become a cornerstone in addressing these challenges (Kumar et al., 2009). Originally developed for taxonomic and biodiversity research, DNA barcoding has been adapted to food science to authenticate raw and processed food products, even when morphological characteristics are degraded or lost (Dawn and Ahn, 2022). The technique has evolved further with the integration of next-generation sequencing (NGS), enabling DNA metabarcoding—a high-throughput approach capable of detecting multiple species in complex mixtures (Dobrovolny et al., 2022). This has proven especially valuable in uncovering adulteration in meat products and composite foods. For example, a 2024 study applying metabarcoding to EU meat samples revealed numerous undeclared animal and plant species, providing evidence of hidden substitutions and labelling violations (Mottola et al., 2024). However, the field is not without its limitations. A 2024 systematic review of metabarcoding in animal-origin food authentication highlighted persistent issues such as inconsistent primer sets, platform-dependent biases, and the need for standardized quality control procedures before the method can be widely adopted for regulatory use (Giusti et al., 2024). In the seafood industry, DNA barcoding continues to reveal significant mislabelling — for instance, a 2021 Italian border-inspection study found 22.5 % mislabelling in imported seafood, with cephalopods mislabelled at 43.8 % and fish at 14.0 % — sometimes involving endangered or protected species (Filonzi et al., 2023; Guardone et al., 2021). The economic impact of food fraud remains uncertain, with global estimates ranging between \$ 10 billion (Robson et al., 2020) to at least \$ 65 billion (Agetu, 2020). In 2017 alone, authorities in Europe seized approximately 9,800 tonnes of non-authentic food (Kendall et al., 2019). These findings illustrate the global scale of the problem and the importance of molecular tools in enforcing food law and protecting endangered species.

This review provides a comprehensive overview of DNA barcoding and its technological advancements, with a focus on its role in ensuring food authenticity and traceability “from gene to table” (Liu K et al., 2022). Current applications

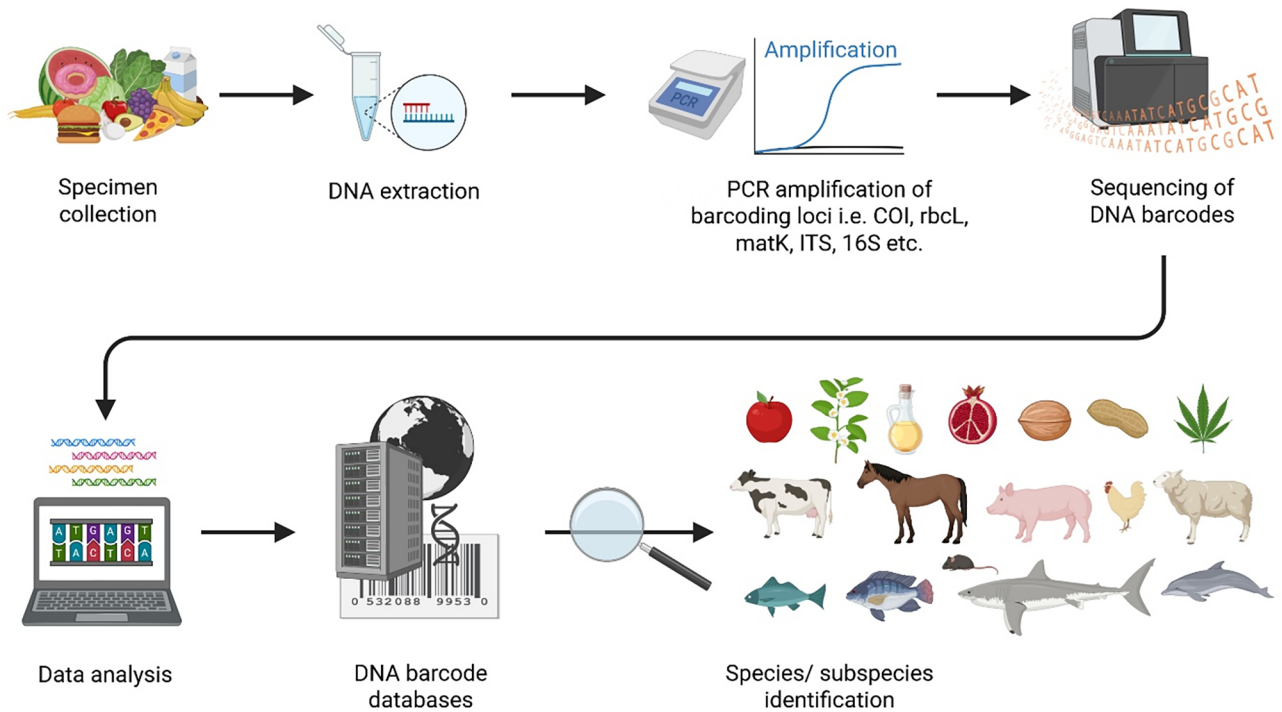
across food categories are reviewed, regulatory perspectives are addressed, and emerging trends including portable sequencing and the integration with blockchain technology are examined (Singh and Sharma, 2023).

## 2. Principles and technological advancements in DNA barcoding

The selection of DNA barcoding markers depends on the target organism. For animals, the cytochrome c oxidase I (COI) remains the standard. In plants, mitochondrial genes evolve too slow, so plastid genes like *rbcL* and *matK* are used instead (CBOL Plant Working Group, 2009). For fungi, the internal transcribed spacer (ITS) region offers high discriminatory power (Schoch et al., 2012).

The barcoding workflow typically begins with sample collection from food matrices (Figure 1). DNA is then extracted using commercial kits or CTAB-based protocols designed to overcome food-specific inhibitors like fats or tannins. Target regions are amplified via PCR, often using nested or multiplex strategies in complex mixtures (Handy et al., 2011). Sequencing is performed through either traditional Sanger methods or whole-genome sequencing. Traditional Sanger sequencing, while reliable, is limited by low throughput and an inability to resolve multiple species in complex mixtures. These constraints have been overcome by the introduction of next-generation sequencing (NGS) technologies. This enables DNA metabarcoding—simultaneous identification of multiple species within a single mixed sample—making the technique particularly valuable for testing multi-ingredient and processed foods like sausages, herbal blends, and convenience meals.

In cases where DNA is degraded—such as in canned or heavily processed foods—shorter “mini-barcodes” (less than 200 base pairs) improve amplification success (Stahl et al., 2023). Mini-barcoding achieved identification rates above 90% across a range of difficult food matrices (Stahl et al., 2023). Nevertheless, the shorter length of mini-barcodes may limit their ability to discriminate between closely related species, and issues such as misidentified specimens, sample confusion, or contamination can further compromise accuracy. However, advances in reference databases and bioinformatic methods have improved reliability, and combining multiple loci or integrating mini-barcodes with full-length markers allows robust species identification (Galimberti et al., 2013; Cheng et al., 2023).



**Figure 1.** The principles of the DNA- barcoding technique (Created in BioRender. Ruppitsch W. (2025) <https://BioRender.com/bwtr3ev>)»

The effectiveness of these tools depends heavily on the quality of the reference databases used. Customized regional or institutional databases are increasingly being incorporated into barcoding workflows to complement public repositories and improve identification accuracy. Such tailored datasets, for example those developed for mammalian and poultry species authentication, allow laboratories to use relevant barcode sequences adapted to their validation needs (Dobrovolsky *et al.*, 2022). Two of the most common global databases are the Barcode of Life Data System (BOLD), which is curated specifically for barcoding and linked to voucher specimens (Ratnasingham and Hebert, 2007), and GenBank, which offers broader coverage but is less curated and more prone to misidentifications (Nilsson *et al.*, 2006). To enhance consistency and data interoperability, initiatives such as the FAIR data principles (Findable, Accessible, Interoperable, Reusable) have been introduced. These guidelines aim to improve database standardization and facilitate the integration of barcoding outputs into regulatory and traceability systems (Wilkinson *et al.*, 2016). Hybrid strategies—such as cross-validating GenBank hits with BOLD entries or building in-house curated databases—are becoming increasingly common for ensuring accuracy in critical applications like regulatory inspections and certification.

Collectively, these technological advancements have pushed DNA barcoding from a niche laboratory method into a mainstream, scalable, and field-deployable tool with broad applications in food integrity and safety.

### 3. Applications in food integrity and safety

One of the most pressing concerns in global food markets is species mislabelling and substitution, which remains prevalent in seafood, meat, and plant-based products. For instance, studies in Thailand have shown that over 24% of seafood products were mislabelled, including cases where endangered species like *Thunnus maccoyii* (Southern bluefin tuna) were substituted for more common varieties (Senathipath *et al.*, 2024). DNA-based methods, including metabarcoding and DNA barcoding, have revealed the presence of undeclared animal species in a substantial proportion of meat products, highlighting widespread mislabelling and the utility of these molecular tools for verifying species composition and ensuring food authenticity (Tantunan and Viljoen, 2021; Hellberg *et al.*, 2017). In the plant sector, DNA markers such as *rbcL*, *matK*, and ITS2 have uncovered extensive adulteration in herbal supplements and traditional medicines (Harris *et al.*, 2024; Chen *et al.*, 2022). High-value oils, such

as olive oil from protected regions, can also benefit from DNA-based traceability tools, which help authenticate geographic origin and detect adulteration with lower-quality oils (Agrimonti et al., 2011). DNA barcoding is also instrumental in detecting adulterants and contaminants that pose health risks to consumers. Even trace amounts of undeclared allergens like peanuts, soy, or crustaceans can trigger serious allergic reactions. DNA-based analyses have proven effective in identifying such allergens in processed foods and sauces, as well as banned substances like bushmeat or endangered species in imported products (Staats et al., 2016). Using metabarcoding techniques, numerous instances of undeclared animal DNA have been detected in products labelled as vegan or allergen-free, which can lead to product recalls and stricter regulatory oversight (Giusti et al., 2023; BeVeg, 2023). The method also enables detection of toxic plant species that may be accidentally or fraudulently incorporated into teas, supplements, or traditional remedies—situations where visual inspection would be inadequate (Sucher and Carles, 2008).

Beyond authenticity and allergen control, DNA barcoding plays a growing role in verifying compliance with religious and ethical food certifications. Halal certification, for example, requires the exclusion of any porcine material; barcoding is routinely employed to test for pork DNA in meat, gelatine, and additives (Shabani et al., 2015). Kosher regulations similarly restrict specific ingredients and combinations, and DNA-based tests can detect the presence of shellfish or non-kosher fish in processed foods (Andronache et al., 2025). Even vegan-labelled products are increasingly scrutinized using

metabarcoding to ensure they are free of trace animal DNA, especially in shared production facilities (Srivathsan et al., 2021).

DNA barcoding is expanding into the realm of microbial food safety (Sabater et al., 2021). Although originally developed for multicellular organisms, adaptations using 16S rRNA and ITS rRNA sequencing now allow for the identification of bacterial, fungal, and parasitic contaminants. This approach has been applied to detect key pathogens such as *Salmonella*, *Listeria*, and *E. coli* in meat and dairy products, as well as toxigenic fungi like *Aspergillus* spp. and *Fusarium* spp. in cereals and spices (Muñoz-Martinez et al., 2025; Gonzalez-Escalona et al., 2017), and protozoan contaminants like *Cryptosporidium* spp. and *Giardia* spp. in water and fresh produce. DNA metabarcoding has been applied in Sardinian sheep cheese processing environments, identifying bacterial strains that could contribute to product contamination and spoilage, highlighting the utility of metabarcoding for monitoring microbial risks in cheese production (Giagnoni et al., 2025). Applications of DNA barcoding in food integrity and safety are shown in Table 1.

4. Regulatory and industrial perspective

Food fraud within the European Union is regulated under Regulation (EC) No. 178/2002 of the European Parliament. Despite significant efforts toward its prevention, access to reliable and publicly available information remains limited (Brooks et al., 2021). European mechanisms such as the Rapid Alert System for Food and Feed (RASFF), established in 1979, and the EU Food Fraud Network,

Table 1. Applications of DNA barcoding in food integrity and safety

Application area	Description
Species mislabelling	Identification of incorrectly labelled or substituted species in meat, fish, etc.
Detection of allergens and contaminants	Detection of trace allergens (e.g., peanut, soy) and harmful substances.
Religious and ethical standards	Verification of the absence of prohibited ingredients in Halal, Kosher, and vegan products.
Microbial contamination	Identification of bacteria, fungi, and parasites relevant to food safety.
Authentication of plant-based products	Analysis of teas, supplements, and herbal blends for undeclared or toxic species.
Verification of declared composition	Detection of undeclared or illegal ingredients in complex processed foods.

launched in 2013, have improved data sharing and cross-border collaboration in addressing food fraud (Brooks *et al.*, 2021). However, the complexity of international supply chains, combined with the highly complex nature of many food products, increases the probability that fraud remains undetected (Manning & Monaghan, 2019).

One contributing factor is that the Hazard Analysis and Critical Control Points (HACCP) system, while effective for food safety, was not designed for the detection of fraudulent practices (Kowalska, 2018). To address this gap, complementary frameworks such as Threat Analysis and Critical Control Points (TACCP) and Vulnerability Assessment and Critical Control Points (VACCP) are increasingly being adopted (Brooks *et al.*, 2021). The effectiveness of these systems, however, depends heavily on the application of advanced analytical techniques capable of verifying food authenticity and preventing adulterated products from entering the supply chain (Manning & Soon, 2016; Fox *et al.*, 2018).

Building on this foundation, DNA-based techniques such as DNA barcoding and metabarcoding are increasingly regarded as valuable tools to strengthen global food authenticity and traceability systems. In the EU, the European Food Safety Authority (EFSA) has acknowledged their scientific potential, particularly regarding identity verification of novel foods including plants, animals, fungi, and algae, but has not yet formally endorsed their widespread use (Veveris, 2024). Metabarcoding, for instance, shows particular promise in detecting undeclared species or complex plant compositions, though its regulatory application remains under development by EFSA and EU Member States (Motola *et al.*, 2024; Giusti *et al.*, 2024).

In the United States, the Food and Drug Administration (FDA) has integrated DNA-based methods into its seafood inspections and guidance materials to support regulatory compliance and ensure seafood authenticity (Handy *et al.*, 2011). These methods use standardized DNA sequences, such as those in the Regulatory Fish Encyclopedia, to detect mislabelled or substituted species throughout the supply chain, enhancing traceability, preventing fraud, and supporting consumer protection (Haile *et al.*, 2008). DNA barcoding represents a valuable tool that helps the FDA ensure the safety and integrity of the U.S. food supply (Jones *et al.*, 2013).

Countries like Canada, Australia, and China also apply DNA barcoding in customs, Halal/Kosher certification, and herbal verification (Fathima *et al.*, 2024;

Agbayeva *et al.*, 2021). Seafood processors use DNA barcoding to confirm species at each supply chain step, helping detect mislabelling and thus improve traceability (Pardo *et al.*, 2020). The technology is also being embedded in blockchain platforms for meat and herbal supplements, linking product records to genetic proof (Gorini *et al.*, 2023).

Globally, the Codex Alimentarius Commission has begun incorporating molecular tools into its food authenticity standards, encouraging harmonization. While Codex recognizes the importance of DNA-based methods for detection and identification of species and specific proteins in foods, their use is not yet mandatory, and implementation depends on national authorities (Codex, 2011). So far, FDA and the Joint Research Centre (JRC) of the European Commission have released detailed protocols, and Codex is evaluating molecular methods for inclusion in international guidelines, however standardized and validated protocols are still needed to ensure harmonized application across countries (JRC, 2024; Codex, 2023).

## 5. Challenges and limitations

Despite its advantages, DNA barcoding faces several limitations that hinder its universal application in food authenticity and safety. A major technical barrier is DNA degradation in processed foods. Thermal, chemical, or mechanical treatments often fragment DNA, making standard barcode regions like COI, *rbcL* or ITS difficult to amplify (Meunier *et al.*, 2008). Mini-barcodes (<200 bp) have been developed to overcome this, though they may lack full taxonomic resolution (Shokralla *et al.*, 2015). Another critical issue is the incompleteness and inconsistency of reference databases (Kartzinel *et al.*, 2025). Many food-relevant species are under-represented, especially among plants, fungi, and minor seafood taxa. Misannotations in public databases such as GenBank can result in misidentifications, prompting researchers to cross-reference with BOLD or use curated in-house databases (Ratnasingham and Hebert, 2007).

A lack of global harmonization means results accepted in one region may not be valid in another (Geary *et al.*, 2019). Data sovereignty is also a concern, particularly where genetic resources from indigenous regions are used without consent, raising compliance issues with the Nagoya Protocol (Prathapan and Rajan, 2020). Misidentification due to contaminated or incomplete data can lead to costly

recalls and reputational harm. Costs and infrastructure constraints remain a major hurdle in low- and middle-income countries. Although technologies have improved accessibility, expenses for reagents, limited lab capacity, and lack of trained staff are persistent challenges (Yuan et al., 2025). Additionally, global databases are biased toward species from developed regions, limiting their relevance for foods originating from Africa, Asia, and Latin America. Greater investment in local databases, training, and technology transfer is essential for equitable global adoption (Yuan et al., 2025).

## 6. Future perspectives

DNA barcoding is evolving beyond species identification into a central tool for transparency, adaptability, and sustainability. Its convergence with blockchain, artificial intelligence (AI), and ecosystem monitoring opens new opportunities for traceability and risk management. Blockchain integration offers secure, tamper-proof traceability by link-

ing DNA-verified species data to immutable digital records. Pilot projects in high-value sectors like seafood and organic meat have already connected QR codes on packaging to DNA-based authenticity logs (Gröppel-Klein et al., 2023; FAO, 2022). This model could become standard for certified products like Halal or sustainably sourced goods. AI and machine learning are transforming how barcoding data is processed. These tools automate species classification, detect anomalies, and integrate genetic, geographic, and trade data to predict fraud and supply chain vulnerabilities (Lokan et al., 2024). Barcoding also supports climate-resilient food systems by enabling rapid identification of shifting species and empowering small producers to validate products for competitive markets and it contributes to reduce food waste through improved sorting, labelling, and quality control (Gorini et al., 2023). In this broader role, DNA barcoding is poised to become essential infrastructure for building ethical, transparent, and sustainable food systems worldwide.

**Disclosure Statement:** No potential conflict of interest was reported by the authors.

**Funding:** This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

## References

- Agetu, R. (2020). Food safety verification by block chain; a consumer-focused solution to the global food fraud crisis. <https://doi.org/10.22004/ag.econ.305254>
- Aghayeva, P., Cozzolino, S., Cafasso, D., Ali-Zade, V., Fineschi, S., & Aghayeva, D. (2021). DNA barcoding of native Caucasus herbal plants: potentials and limitations in complex groups and implications for phylogeographic patterns. *Biodiversity Data Journal*, 9, e61333. <https://doi.org/10.3897/BDJ.9.e61333>
- Agrimonti, C., Vietina, M., Pafundo, S., & Marmiroli, N. (2011). The use of food genomics to ensure the traceability of olive oil. *Trends in Food Science & Technology*, 22(5), 237–244. <https://doi.org/10.1016/j.tifs.2011.02.002>
- Andronache, J., Cichna-Markl, M., Dobrovolny, S., & Hochegger, R. (2025). Development of a DNA metabarcoding method for the identification of crustaceans (Malacostraca) and cephalopods (Coleoidea) in processed foods. *Foods*, 14(9), 1549. <https://doi.org/10.3390/foods14091549>
- BeVeg, (2023, January 28). Vegan products recalled for undeclared animal allergens. *BeVeg*. <https://www.beveg.com/vegan-products-recalled-for-undeclared-animal-allergens/>
- Bolyen, E., Rideout, J. R., Dillon, M. R., et al. (2019). Reproducible, interactive, scalable and extensible microbiome data science using QIIME 2. *Nature Biotechnology*, 37(8), 852–857. <https://doi.org/10.1038/s41587-019-0209-9>
- Brooks, C., Parr, L., Smith, J. M., Buchanan, D., Snioch, D., & Hebishy, E. (2021). A review of food fraud and food authenticity across the food supply chain, with an examination of the impact of the COVID-19 pandemic and Brexit on food industry. *Food Control*, 130, 108171. <https://doi.org/10.1016/j.foodcont.2021.108171>
- CBOL Plant Working Group, (2009). A DNA barcode for land plants. *Proceedings of the National Academy of Sciences*, 106(31), 12794–12797. <https://doi.org/10.1073/pnas.0905845106>
- Cheng, X., Liu, S., Zhang, Y., Li, Q., & Wang, J. (2023). The devil is in the details: Problems in DNA barcoding practices indicated by systematic evaluation of insect barcodes. *Frontiers in Ecology and Evolution*, 11, 1149839. <https://doi.org/10.3389/fevo.2023.1149839>
- Codex Alimentarius Commission, (2010). *Guidelines on performance criteria and validation of methods for detection, identification and quantification of specific DNA sequences and specific proteins in foods (CAC/GL 74-2010)*. Food and Agriculture Organization of the United Nations. [https://www.fao.org/fileadmin/user\\_upload/gmfp/resources/CXG\\_074e.pdf](https://www.fao.org/fileadmin/user_upload/gmfp/resources/CXG_074e.pdf)

- Dawan, J., & Ahn, J. (2022). Application of DNA barcoding for ensuring food safety and quality. *Food Science and Biotechnology*, 31(11), 1355–1364. <https://doi.org/10.1007/s10068-022-01143-7>
- Dobrovolny, S., Uhlig, S., Frost, K., Schlierf, A., Nichani, K., Simon, K., Cichna-Markl, M., & Hohegger, R. (2022). Interlaboratory validation of a DNA metabarcoding assay for mammalian and poultry species to detect food adulteration. *Foods*, 11(8), 1108. <https://doi.org/10.3390/foods11081108>
- Dobrovolny, S., Uhlig, S., Frost, K., Schlierf, A., Nichani, K., Simon, K., Cichna-Markl, M., & Hohegger, R. (2022). Interlaboratory validation of a DNA metabarcoding assay for mammalian and poultry species to detect food adulteration. *Foods*, 11(8), 1108. <https://doi.org/10.3390/foods11081108>
- European Commission Joint Research Centre, (2024, November 29). New JRC research helps expose food fraud. *EU Science Hub*. [https://joint-research-centre.ec.europa.eu/jrc-news-and-updates/new-jrc-research-helps-expose-food-fraud-2024-11-29\\_en](https://joint-research-centre.ec.europa.eu/jrc-news-and-updates/new-jrc-research-helps-expose-food-fraud-2024-11-29_en)
- Fathima, A. M., Rahmawati, L., Windarsih, A., & Suratno. (2024). Advanced halal authentication methods and technology for addressing non-compliance concerns in halal meat and meat products supply chain: A review. *Food Science of Animal Resources*, 44(6), 1195–1212. <https://doi.org/10.5851/kosfa.2024.e75>
- Filonzi, L., Ardenghi, A., Rontani, P. M., Voccia, A., Ferrari, C., Papa, R., Bellin, N., & Nonnis Marzano, F. (2023). Molecular Barcoding: A Tool to Guarantee Correct Seafood Labelling and Quality and Preserve the Conservation of Endangered Species. *Foods* (Basel, Switzerland), 12(12), 2420. <https://doi.org/10.3390/foods12122420>
- Food and Agriculture Organization of the United Nations & World Health Organization, (2023). *Codex Alimentarius Commission procedural manual* (28th ed.). FAO/WHO. <https://www.fsai.ie/getattachment/50461620-bbe9-4891-b6b6-4aec6aa2a11f/final-the-suitability-of-analytical-methods-for-assessing-food-authenticity.pdf>
- Food and Agriculture Organization of the United Nations, (2022). *Blockchain application in seafood value chains* (FAO Fisheries and Aquaculture Circular No. 1257). FAO. <https://openknowledge.fao.org/server/api/core/bitstreams/e7401fd1-ee65-4523-9f42-abf874605427/content>
- Fox, M., Mitchell, M., Dean, M., et al. (2018). The seafood supply chain from a fraudulent perspective. *Food Security*, 10, 939–963. <https://doi.org/10.1007/s12571-018-0826-z>
- Galimberti, A., de Mattia, F., Losa, A., Bruni, I., Federici, S., Casiraghi, M., Martellos, S., & Labra, M. (2013). DNA barcoding as a new tool for food traceability. *Food Research International*, 50(1), 55–63. <https://doi.org/10.1016/j.foodres.2012.09.036>
- Geary, J., Reay, T., & Bubela, T. (2019). The impact of heterogeneity in a global knowledge commons: Implications for governance of the DNA barcode commons. *International Journal of the Commons*, 13(2), 909–930. <https://doi.org/10.5334/ijc.861>
- Giagnoni, L., Salza, S., Tondello, A., Zardinoni, G., Saptarathi, D., Piras, G., Melillo, R., Cecchinato, A., Stevanato, P., Tedde, T., Mudadu, A. G., Squartini, A., & Spanu, C. (2025). DNA metabarcoding for the identification and relative abundance assessment of general and potentially pathogenic bacteria in Sardinian sheep cheese processing environments. *International Journal of Food Microbiology*, 442, 111353. <https://doi.org/10.1016/j.ijfoodmicro.2025.111353>
- Giusti, A., Malloggi, C., Magagna, G., Filipello, V., & Armani, A. (2023). Is the metabarcoding ripe enough to be applied to the authentication of foodstuff of animal origin? A systematic review. *Comprehensive Reviews in Food Science and Food Safety*, 23(1), e13256. <https://doi.org/10.1111/1541-4337.13256>
- Gonzalez-Escalona, N., Hammack, T. S., & Monday, S. R. (2017). High-throughput sequencing for detection of foodborne pathogens. *Frontiers in Microbiology*, 8, 2029. <https://doi.org/10.3389/fmicb.2017.02029>
- Gorini, T., Mezzasalma, V., Deligia, M., De Mattia, F., Campone, L., Labra, M., & Frigerio, J. (2023). Check Your Shopping Cart: DNA Barcoding and Mini-Barcoding for Food Authentication. *Foods*, 12(12), 2392. <https://doi.org/10.3390/foods12122392>
- Gröppel-Klein, A., & Kirsch, B. (2023). Design of cues on supply chain encryption through blockchain: Effects on consumer trust in packaged “ecologically produced” meat. *Marketing ZPF – Journal of Research and Management*, 45(3), 38–52. [doi.org/10.15358/0344-1369-2023-3-38](https://doi.org/10.15358/0344-1369-2023-3-38)
- Guardone, L., Tinacci, L., Costanzo, F., Azzarelli, D., D’Amico, P., Tasselli, G., Magni, A., Guidi, A., Nucera, D. M., & Armani, A. (2017). DNA barcoding as a tool for detecting mislabeling on incoming fishery products from third countries: An official survey conducted at the Border Inspection Post of Livorno–Pisa (Italy). *Food Control*, 80, 204–216. <https://doi.org/10.1016/j.foodcont.2017.03.056>
- Haile, F. Y., Zemlak, T., Mason, J. A., & Hebert, P. D. N. (2008). Potential use of DNA barcodes in regulatory science: Applications of the Regulatory Fish Encyclopedia. *Journal of Food Protection*, 71(1), 210–217. <https://doi.org/10.4315/0362-028X-71.1.210>
- Handy, S. M., Deeds, J. R., Ivanova, N. V., Hebert, P. D., Hanner, R. H., Ormos, A., Weigt, L. A., Moore, M. M., & Yancy, H. F. (2011). A single-laboratory validated method for the generation of DNA barcodes for the identification of fish for regulatory compliance. *Journal of AOAC International*, 94(1), 201–210.
- Harris, C. M., Kim, D. Y., Jordan, C. R., Miranda, M. I., & Hellberg, R. S. (2024). DNA barcoding of herbal supplements on the US commercial market associated with COVID-19. *Phytochemical Analysis*, 35(4), 664–677. <https://doi.org/10.1002/pca.3320>
- Harris, C. M., Kim, D. Y., Jordan, C. R., Miranda, M. I., & Hellberg, R. S. (2024). DNA barcoding of herbal supplements on the US commercial market associated with the purported treatment of COVID-19. *Phytochemical Analysis*, 35(4), 664–677. <https://doi.org/10.1002/pca.3320>
- Hebert, P. D. N., Cywinska, A., Ball, S. L., & deWaard, J. R. (2003). Biological identifications through DNA barcodes. *Proceedings of the Royal Society B: Biological Sciences*, 270(1512), 313–321. <https://doi.org/10.1098/rspb.2002.2218>
- Hebert, P. D., Cywinska, A., Ball, S. L., & deWaard, J. R. (2003). Biological identifications through DNA barcodes. *Proceedings. Biological Sciences*, 270(1512), 313–321. <https://doi.org/10.1098/rspb.2002.2218>

- Hellberg, R. S., Hernandez, B. C., & Hernandez, E. L. (2017). Identification of meat and poultry species in food products using DNA barcoding. *Food Control*, 80, 23–28. <https://doi.org/10.1016/j.foodcont.2017.04.025>
- Jones, Y. L., Peters, S. M., Weland, C., Ivanova, N. V., & Yancy, H. F. (2013). Potential use of DNA barcodes in regulatory science: Identification of the U.S. Food and Drug Administration's "Dirty 22," contributors to the spread of foodborne pathogens. *Journal of Food Protection*, 76(1), 144–149. <https://doi.org/10.4315/0362-028X.JFP-12-168>
- Kartzinel, T. R., Hoff, H. K., Divoll, T. J., et al. (2025). Global availability of plant DNA barcodes as genomic resources to support basic and policy-relevant biodiversity research. *Molecular Ecology*, 34(7), e17712. <https://doi.org/10.1111/mec.17712>
- Katerinopoulou, K., Kontogeorgos, A., Salmas, C. E., Patakas, A., & Ladavos, A. (2020). Geographical Origin Authentication of Agri-Food Products: A Review. *Foods* (Basel, Switzerland), 9(4), 489. <https://doi.org/10.3390/foods9040489>
- Kendall, H., Clark, B., Rhymer, C., Kuznesof, S., Hajslova, J., Tomaniova, M., Brereton, P., & Frewer, L. (2019). A systematic review of consumer perceptions of food fraud and authenticity: A European perspective. *Trends in Food Science & Technology*, 94, 79–90. <https://doi.org/10.1016/j.tifs.2019.09.014>
- Kowalska, A. (2018). The Study of the Intersection Between Food Fraud/Adulteration and Authenticity. *Acta Universitatis Agriculturae et Silviculturae Mendelianae Brunensis*, 66(5), 1275–1286. doi: 10.11118/actaun201866051275
- Kumar, P., Gupta, V. K., Misra, A. K., Modi, D. R., & Pandey, B. K. (2009). Potential of molecular markers in plant biotechnology. *Plant Omics*, 2(4), 141–162.
- Li, Y. C., Liu, S. Y., Meng, F. B., Liu, D. Y., Zhang, Y., Wang, W., & Zhang, J. M. (2020). Comparative review and the recent progress in detection technologies of meat product adulteration. *Comprehensive Reviews in Food Science and Food Safety*, 19(4), 2256–2296. <https://doi.org/10.1111/1541-4337.12579>
- Liu, K., Xing, R., Sun, R., Ge, Y., & Chen, Y. (2022). An accurate and rapid way for identifying food geographical origin and authenticity: Editable DNA-traceable barcode. *Foods*, 12(1), 17. <https://doi.org/10.3390/foods12010017>
- Lokanan, M. E., & Maddhesia, V. (2024). Supply chain fraud prediction with machine learning and artificial intelligence. *International Journal of Production Research*, 63(1), 286–313. <https://doi.org/10.1080/00207543.2024.2361434>
- Manning, L., & Monaghan, J. (2019). Integrity in the fresh produce supply chain: solutions and approaches to an emerging issue. *The Journal of Horticultural Science and Biotechnology*, 94(4), 413–421. <https://doi.org/10.1080/14620316.2019.1574613>
- Manning, L., & Soon, J. M. (2016). Food safety, food fraud, and food defense: A fast evolving literature. *Journal of Food Science*, 81(4), R823–R834. <https://doi.org/10.1111/1750-3841.13256>
- Meusnier, I., Singer, G. A. C., Landry, J. F., Hickey, D. A., Hebert, P. D. N., & Hajibabaei, M. (2008). A universal DNA mini-barcode for biodiversity analysis. *BMC Genomics*, 9, 214. <https://doi.org/10.1186/1471-2164-9-214>
- Mottola, A., Intermite, C., Piredda, R., Lorusso, L., Rannieri, L., Carpino, S., Celano, G. V., & Di Pinto, A. (2024). DNA metabarcoding approach as a potential tool for supporting official food control programs: A case study. *Foods*, 13(18), 2941. <https://doi.org/10.3390/foods13182941>
- Muñoz-Martínez, T. I., Rodríguez-Hernández, B., Rodríguez-Montaño, M., Alfau, J., Reyes, C., Fernandez, Y., ... Maroto-Martín, L. O. (2025). Unlocking the hidden microbiome of food: The role of metagenomics in analyzing fresh produce, poultry, and meat. *Applied Microbiology*, 5(1), 26. <https://doi.org/10.3390/applmicrobiol5010026>
- Nilsson, R. H., Ryberg, M., Kristiansson, E., Abarenkov, K., Larsson, K. H., & Kõljalg, U. (2006). Taxonomic reliability of DNA sequences in public sequence databases: A fungal perspective. *PLOS ONE*, 1(1), e59. <https://doi.org/10.1371/journal.pone.0000059>
- Ortea, I., Pascoal, A., Cañas, B., Gallardo, J. M., Barros-Velázquez, J., & Calo-Mata, P. (2012). Food authentication of commercially-relevant shrimp and prawn species: From classical methods to foodomics. *Electrophoresis*, 33(15), 2201–2211. <https://doi.org/10.1002/elps.201100576>
- Pardo, M. Á., & Jiménez, E. (2020). DNA barcoding revealing seafood mislabeling in food services from Spain. *Journal of Food Composition and Analysis*, 91, 103521. <https://doi.org/10.1016/j.jfca.2020.103521>
- Prathapan, K. D., & Rajan, P. D. (2020). Submission on Article 10 of the Nagoya Protocol: Peer-review comments on benefit-sharing from digital sequence information on genetic resources. *Convention on Biological Diversity*. <https://www.cbd.int/abs/Art-10/Peer-review/Prathapan-Rajan.pdf>
- Ratnasingham, S., & Hebert, P. D. N. (2007). BOLD: The Barcode of Life Data System. *Molecular Ecology Notes*, 7(3), 355–364. <https://doi.org/10.1111/j.1471-8286.2007.01678.x>
- Sabater, C., Cobo-Díaz, J. F., Álvarez-Ordóñez, A., Ruas-Madiedo, P., Ruiz, L., & Margolles, A. (2021). Novel methods of microbiome analysis in the food industry. *International Microbiology*, 24(4), 593–605. <https://doi.org/10.1007/s10123-021-00215-8>
- Schoch, C. L., Seifert, K. A., Huhndorf, S., Robert, V., Spouge, J. L., Levesque, C. A., Chen, W., & Fungal Barcoding Consortium. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for fungi. *Proceedings of the National Academy of Sciences*, 109(16), 6241–6246. <https://doi.org/10.1073/pnas.1117018109>
- Senathipathi, D. N., Benjakul, S., & Sukkapat, P. (2024). DNA barcoding revealed mislabeling of imported seafood products in Thailand. *Fishes*, 9(6), 215. <https://doi.org/10.3390/fishes9060215>
- Shabani, H., Mehdizadeh, M., Mousavi, S. M., Robotjazi, E., Rastegar, H., & Alebouyeh, M. (2015). Halal authenticity of gelatin using PCR. *Food Chemistry*, 184, 203–206. <https://doi.org/10.1016/j.foodchem.2015.02.140>
- Shokralla, S., Hellberg, R. S., Handy, S. M., King, I., & Hajibabaei, M. (2015). A DNA mini-barcoding system for authentication of processed fish products. *Scientific Reports*, 5, 15894. <https://doi.org/10.1038/srep15894>

- Singh, V., & Sharma, S. K. (2023).** Application of blockchain technology in shaping the future of food industry based on transparency and consumer trust. *Journal of food science and technology*, 60(4), 1237–1254. <https://doi.org/10.1007/s13197-022-05360-0>
- Srivathsan, A., Sha, J., Vogler, A. P., & Meier, R. (2021).** Metabarcoding to detect animal DNA in vegan products. *Methods in Ecology and Evolution*, 12(7), 1443–1451. <https://doi.org/10.1111/2041-210X.13619>
- Staats, M., Arulandhu, A. J., Gravendeel, B., Holst-Jensen, A., & Scholtens, I. (2016).** Advances in DNA metabarcoding for food and wildlife forensic species identification. *Analytical and Bioanalytical Chemistry*, 408(19), 4615–4630. <https://doi.org/10.1007/s00216-016-9595-8>
- Stamatis, C., Sarri, C.A., Moutou, K.A., Argyrakoulis, N., Galara, I., Godosopoulos, V., Kolovos, M., Liakou, C., Stasinou, V., & Mamuris, Z. (2015).** What do we think we eat? Single tracing method across foodstuff of animal origin found in Greek market. *Food Research International*, 69, 151–155. <https://doi.org/10.1016/j.foodres.2014.12.033>
- Sucher, N. J., & Carles, M. C. (2008).** Genome-based approaches to the authentication of medicinal plants. *Planta Medica*, 74(6), 603–623. <https://doi.org/10.1055/s-2008-1074517>
- Tantuan, S. S., & Viljoen, C. D. (2021).** Determining the presence of undeclared animal species using real-time PCR in canned and ready-to-eat meat products in South Africa. *Journal of Food Science and Technology*, 58(7), 2699–2704. <https://doi.org/10.1007/s13197-020-04776-w>
- U.S. Food and Drug Administration, (2023).** *Seafood list guidance documents and regulatory information*. <https://www.fda.gov/food/seafood-guidance-documents-regulatory-information/fda-seafood-list-updates-2023>
- Ververis, E. (2024).** *EFSA novel food guidance update*. European Food Safety Authority. <https://www.efsa.europa.eu/sites/default/files/2024-03/ermolaos-ververis-novel-food-guidance-update.pdf>
- Wilkinson, M. D., Dumontier, M., Aalbersberg, I. J., Appleton, G., Axton, M., Baak, A., Blomberg, N., Boiten, J. W., da Silva Santos, L. B., Bourne, P. E., Bouwman, J., Brookes, A. J., Clark, T., Crosas, M., Dillo, I., Dumon, O., Edmunds, S., Evelo, C. T., Finkers, R., ... Mons, B. (2016).** The FAIR guiding principles for scientific data management and stewardship. *Scientific Data*, 3, 160018. <https://doi.org/10.1038/sdata.2016.18>
- Yao, H., Song, J., Liu, C., Luo, K., Han, J., Li, Y., Pang, X., Xu, H., Zhu, Y., Xiao, P., Chen, S., & Leon, C. (2010).** Use of ITS2 region as the universal DNA barcode for plants and animals. *PLOS ONE*, 5(10), e13102. <https://doi.org/10.1371/journal.pone.0013102>
- Yuan, H., Han, J., Yang, M., Chen, S., & Pang, X. (2025).** DNA metabarcoding: Current applications and challenges. *Journal of Agricultural and Food Chemistry*, 73(33), 20616–20632. <https://doi.org/10.1021/acs.jafc.5c06002>

#### Authors info

**Aleksandra Martinović**, <https://orcid.org/0000-0002-3177-9868>

**Nada Raičević**, <https://orcid.org/0009-0007-8354-9507>

**Beatriz Daza Prieto**, <https://orcid.org/0000-0002-7138-5441>

**Werner Ruppitsch**, <https://orcid.org/0000-0001-9940-3333>

**Marija Vugdelić**, <https://orcid.org/0000-0001-5638-3942>